

Animal models for acquired heterotopic ossification

Joris Anthonissen, Christian Ossendorf, Ulrike Ritz, Alexander Hofmann, Pol Maria Rommens

From the Department of Orthopaedics and Trauma Surgery, University of Mainz, Germany

Heterotopic ossification (HO), the ectopic formation of bone in soft tissues, is a relevant musculoskeletal disorder that, by reduction of range of motion, may lead to significant impairment of quality of live. HO can either be acquired or hereditary. Acquired HO is seen most often after hip prosthetic surgery and pelvic trauma. In contrast, hereditary HO is commonly observed in the axial skeleton, but can affect every joint. Substantial effort has been directed towards understanding the pathophysiology and towards finding both, effective prophylactic and therapeutic treatments. Every improvement of the understanding of the pathophysiologic changes underlying HO as well as the rationale of prophylactic and therapeutic treatment regimens in the end, is based on the study of appropriate animal models. Although intriguing models of 'genetic' HO have been developed recently, their relevance to acquired HO remains questionable.

As there is still neither proper treatment nor reliable prophylaxis, animal models will remain important in the study of HO. Currently, there are 6 different animal models regularly used for the study of acquired HO. Some of these models can reflect a merely particular part of the disease. Hence, selection of the appropriate animal model for the study of HO is exceedingly important. The present paper reviews the history and major features of the different animal models of acquired HO, and reveals some of the insights gained through the study of animal models; important biochemical and pathophysiological key features are highlighted. Clinical studies have proved indometacine, celecoxib and radiation therapy to be effective in reducing the occurrence of HO, but not always be able to prevent it.

INTRODUCTION

The first description of heterotopic ossification (HO) goes back to a scientific contribution by Patin in children suffering from 'myositis ossificans progressiva' published in 1692 (68). Later, a more distinct description of HO was provided by Riedel *et al*, as well as by Dejerine and Ceilier in 1883 and 1918, respectively (16,24,75). Although HO is usually defined as a new formation of trabecular bone including bone marrow in soft-tissues, where bone usually does not occur (4,18), there is no consensus on the name, definition and classification of HO (64,103).

HO is no trivial disease ; the formation of heterotopic bone can lead to a limitation of the range of motion and may have serious consequences for the quality of life of patients (*103*).

- Joris Anthonissen, Resident.
- Christian Ossendorf, MD, MSc.
- Ulrike Ritz, PhD.
- Alexander Hofmann, MD.
- Pol Maria Rommens, MD.

Department of Orthopaedics and Trauma Surgery, University of Mainz, Germany.

Corresponding : Joris Anthonissen.

E-Mail : joris.anthonissen@unimedizin-mainz.de.

Mailing adress: Department of Orthopaedics and Trauma Surgery, University of Mainz, Germany, Langenbeckstraße 1 55131 Mainz, Germany.

© 2014, Acta Orthopædica Belgica.

Two types of HO are distinguished : genetic and acquired (87). Many authors suggest a similar pathogenesis of genetically caused and acquired HO (1, 41,55,86,90). Genetically caused HO consists mainly of two disorders, fibrodysplasia ossificans progressiva (FOP) (43) and progressive osseous heteroplasia (POH). Vast deposits of heterotopic bone characterize these two entities with progressive accumulation around several joints. This process eventually leads to severe disability and early death due to pneumonia (56). A wealth of research has recently been done on the pathophysiology of FOP (42,110, 111). The genetic regulation of some bone morphogenetic proteins (BMP) and its inhibitory factors were proven to be impaired, and suggested as a major effector underlying genetically caused HO (42,110,111). In contrast, the acquired form usually is either precipitated by trauma or has a neurogenic cause (87). HO may occur after virtually any type of musculoskeletal trauma. The most common site for the formation of HO is the pelvic bone after open-reduction internal-fixation (ORIF) for acetabular fracture, followed by the hip after total hip arthroplasty (THA) (2,4,8,27,77,78,95); but also after orthopaedic procedures of the knee, shoulder or elbow, and fractures, respectively. Even after joint dislocation or direct soft tissue trauma, the formation of HO was observed (22,30,36). In contrast to the hip and pelvis, abdominal incisions, wounds (specially war amputation wounds), the kidneys and the gastrointestinal tracts are less commonly encountered sites of posttraumatic HO (20,32,37,65,71,72). The other form of acquired HO (i.e. HO with neurogenic cause) occurs after injury to the nervous system, usually after traumatic brain injury or spinal cord injury (20,67). Bone formation following neurologic injury tends to form in para-articular sites. The most commonly affected joint in neurogenic HO is the hip, followed by shoulder and elbow. Neurogenic HO rarely occurs around the knee (21,38).

The incidence of HO after THA ranges between 16 and 53%, (103) but only a minority of the patients becomes symptomatic (3-7%) (4,20). In contrast, the incidence of HO following neurologic trauma is reported to be 10-30% (92). In the prevention of HO after THA, in patients with high risk, indomethacin, celecoxibs and radiation are generally accepted as

the treatment of choice (4,76,103). In the prevention of HO after neurologic trauma, much controversy exists on the use of range-of-motion exercises (44, 53). Therapeutic options in HO are limited and a high recurrence rate is observed. Currently the most common treatment is surgical resection and radiation therapy to prevent recurrence of HO (92).

As the pathophysiology of HO remains unclear, the limited current available prophylactic and therapeutic interventions appear to be neither sophisticated nor always effective. Practically every better understanding of the pathophysiologic changes underlying HO and the rationale of prophylactic and therapeutic treatment regimens owe their origins to the study of animal models. Animal models reflecting pathophysiology, prevention and treatment will thus play an important role in the future (4). Currently 6 major animal models are used. Here, we review their history and major features including the respective advantages and disadvantages. The most important insights gained through the study of these animal models are highlighted as well.

ANIMAL MODELS

Achilles tenotomy model

The occurrence of HO in humans after achilles tenotomy was first described by Jones in 1932 (39). He observed painful ossifications in achilles tendons 10 years or more after achilles tenotomy. This observation presumably ended in the development of an achilles tenotomy model in rats in 1953 (9). Buck described a simple mid-way division of the achilles tendon using a sharp razor blade after skin incision and blunt dissection. Thereafter, only the skin was closed, i.e. no adaptation of the tendon ends using sutures. In this model, HO could be shown in all specimens by the end of a three-month-period (81).

In 1969, Salah was able to show that it isn't necessary to divide the achilles tendon. He observed that the squeezing of the tendon with an artery forceps also led to HO. Furthermore, he demonstrated that, when two ligatures are placed around the achilles tendon, the segment between the ligatures is gradually converted into a large ossicle (82). He also proved that no HO is formed when the calf muscles are denervated. Therefore the pull of the muscles on the achilles tendon seemed to be of great relevance. In 1983, McClure wrote an article that is judged as the standard work on the Achilles tenotomy model. He applied the achilles surgery described by Buck to mice and found that ectopic bone developed in 60% of animals after 5 weeks and in 100% after 10 weeks (57). The advantages of this model are its relative simplicity and excellent predictability. However, the molecular mechanisms of HO induced by Achilles tenotomy are poorly understood, and the relevance to clinical conditions is unclear since ectopic bone formation in the achilles tendon is a rare condition in humans (40). Recently this model is often used in rats and mice to research different preventive strategies to reduce the occurrence of HO formation (12,51,83,114,116). The validity of these finding in comparison to humans is however largely unclear. Also in the pathogenetic research of HO this model is used with the same limitations (40.52).

Immobilisation-manipulation (Michelsson) model

In 1980, Michelsson et al could show that the repeated and intensified mobilization of the knee joint in rabbits causes formation of HO in (rabbit) quadriceps muscles (58). Michelsson developed a model that was characterized by rabbit's knee immobilisation for 5 weeks with a plastic splint and elastic bands. During this period, the splint was removed each day and the knee was passively and intensively mobilized for 5 minutes through the full range of motion. After this 5-week period, the splint was removed and animals were allowed to move freely. During this experiment, HO was seen radiographically in all animals at the end of the 5-week period. Further growth of HO could be demonstrated after the 5 first weeks. In a second publication in 1994, Michelson et al showed, by placing a membrane between the quadriceps and the femur, that isolation of bone from muscles prevented the development of experimental callus-like HO (59). Therefore, Hardy criticized in 1997 the finding of Michelson arguing that it is not HO that is seen in his model but a dystrophic calcification

(33) In a reaction on this Michelson stated that bone developed in the vastus intermedius muscle where in normal circumstances no bone is found and that therefore his findings should be called HO. This discussion highlights the problems that arise from the lack of a consensus on the definition of HO. Since his publication in 1980 several authors have used this model to study the development and prevention of HO in rabbits (5,60,98,104,105). The first sign of osteoblastic activity was seen in the periosteum, and the new bone was often formed in continuity with the periosteum. Interestingly, early changes in prostaglandins preceded bone formation consistent with the hypothesis that inflammation is the basis of the heterotopic bone formation in that process (98). Although it seems that the interaction between the periosteum and the necrotic muscle are necessary for the formation of HO, since the introduction of a plastic membrane between bone and muscle prevents bone formation (59), the precise inductive stimulus has not been identified in this model. Therefore its relevance to human HO remains unclear.

Implantation / injection models

The most commonly used animal models in the research of possible therapy and prevention of HO involve the surgical implantation of BMP containing matrices or injection of BMP containing substances at heterotopic sites (40). These models have been employed for over 80 years. In 1938, Levander was able to induce the formation of cartilage and bone in 23% of animals by injecting alcoholic extracts of autologous bone into the rectus femoris muscle of rabbits (3,48,49). Bertelsen (1940) proved that alcoholic extracts of bone marrow (83%) were superior to extracts of cortex, epyphisis and periosteum (48%) (6). Lacroix (1945) obtained bone with hematopoietic marrow with extracts of epiphyses of new-born rabbits (45). He suggested that the hypothetically inducing substance be called 'osteogenin'. Lagos (1946) and Heinen (1949) challenged these findings by stating that the injection of alcohol alone had the same effect as injecting an alcoholic bone extract (35,46). They therefore denied the existence of a specific osteogenic substance. In 1957 Danis found that after heterotopic transplantation of bone marrow, bone was formed at the site of heterotopic implantation (15). Chalmers, Burnwell and Friedenstein confirmed these findings in 1959, 1964 and 1966, respectively (10,11,19). In 1965 Urist showed in different animals that samples of diaphyseal decalcified bone implanted in a pouch in the belly of a muscle gave rise to new bone formation by what he called auto-induction (100). This model of implanting demineralised bone matrix into soft tissue was rapidly adopted by others and is still used to research the induction of bone / HO in soft tissue and how to prevent the formation of HO (11,14,17,54,61,97,117). Reddi (1972) characterised this bone formation extensively and found that it mirrors the normal process of in vivo cartilage and bone formation (74). Urist and collaborators identified in 1979 the active component in the bone extracts used in these early experiments and named it bone morphogenetic protein (BMP) (101, 102). Wozney et al were able to repeat this experiment using partially purified BMP proteins (108). These experiments proved the existence of a specific osteogenic substance that Lacroix called osteogenin (45). Currently, the most widely used approach is BMP-matrigel implantation at heterotopic sides (25). Many modifications/variations of this method have been used in different species under different conditions to assess the pathogenesis and prevention of HO (31,34,47,62,88,89,106). In recent experiments, transfection of BMP coding genes into animal soft tissue is used to get a better knowledge of the pathogenesis of HO and bone formation in general. In these experiments adenoviruses, retroviral viruses, and plasmid particles containing BMP-genes (mostly BMP 2 or 4) are used to induce HO (26,66,69,70,109). Inhibitors of bone formation as Noggin and Gremlin are also injected in this way to evaluate their function (23,99,115).

Another intriguing version of this model researches the osteoinductive ability of certain biomaterials, such as micro-porous calcium phosphate ceramic particles, that do not release BMP or other known osteogenic factors (63). The mechanism of osteoinduction by such biomaterials is not currently clear, although the geometry of the material is thought to play an important role (40,112). Generally, heterotopic implantation / injection models are straightforward, reliable, and mechanistically relevant to human HO. However, certain limitations do exist : (1) they are artificial systems that may create non-physiologically high local concentrations of osteogenic factors at implanted sites leading to effects not relevant to the human disorder, (2) the implantation is a local event and thus has limited ability to mimic the potential effects of the involvement of systemic factors.

Hip surgery (Schneider) model

Schneider *et al* described a model to simulate the pathogenesis of HO after hip arthroplasty in 1998 (84). In this model, male New Zealand rabbits were treated similarly to human hip arthroplasty using a standard approach anterolateral to the hip. The left hip of the animals underwent muscle injury by clamping to produce ischemia of gluteus maximus and medius muscles. The right hip underwent no muscle injury and served as a control. The medullary canal of the femur was opened and reamed comparable to the implantation of a hip prosthesis. The reaming-debris was left in situ. This straightforward model was reported to produce HO with high reliability in 17 of 18 animals with no significant difference in amounts of bone formation between muscle injury side and control side.

Rumi successfully used this model in 2005 to assess the optimal timing of prophylactic preoperative radiation and to identify the origin of osteoprogenitor cells responsible for HO (79,80).

Toom *et al* used this model as described by Schneider in rats to research the role of osteoprogenitor cells from the femoral canal (96). In their research, the induction of HO (without BMP-2 implants) was not successful. Only cartilage was found in the examined samples. This is probably because the animals were sacrificed at 3 and 21 days already. In contrast, as described in the original publication, Schneider *et al* studied the animals radiographically after 1 to 7 months and histologically after 7 months only (84). Using this hip surgery model, Tannous *et al* proved that the formation of bone in HO is endochondral. In this experiment the first 21 days only cartilage was formed and only later this cartilage would calcify and reorganise to lamellar bone (94). This model is a mechanism-based model, reflecting a known cause of HO in humans. It seems therefore a suited model to study the formation of HO after THA. Whether this model is also relevant to other forms and causes of HO remains questionable.

Direct trauma models

Traumatic muscle injury can lead to bone formation in soft tissue of humans. McCarthy and Sundaram (2005) termed this type of bone formation 'myositis ossificans circumscripta' (56). They described it as a self-limiting disorder in which an osseous mass develops close to bones and joints, mostly initiated by a trauma and typically seen in patients 15-30 years of age.

Efforts to establish trauma-induced models had only limited success. Back in 1904, Haga and Fujimura reported to have evoked ossification in traumatized animal tissue (29). Gruber challenged this finding in 1913 by stating that Haga and Fujimura did not reveal their method nor their exact results (28). In line with his criticism, Gruber self failed to induce ossification in rabbit thigh muscle using single hammer strikes. In 1926 Stone was unable to detect ossification in dogs after striking the anterior surface of the thigh during partial or complete relaxation (91). Using a mini version of a pile driver, Zaccalini and Urist (1964) were not able to induce HO in rabbit thigh (113) In an intriguing study, Collins et al (1965) induced bone formation by stripping the periosteum of the thigh bone and damaging the overlying muscle (13). Ossification was enhanced by repeated blunt trauma over the thigh after injury. Walton et al (1983) reported the induction of intramembranous ossification within scar tissue in sheep following blunt trauma of the thigh (107). The induction of ossification was only successful in 16.6% of traumatized thighs, further, intramembranous and not endochondral ossification was the histological feature within scar tissue. More recently Tannous et al induced heterotopic ossification in rat in an extremity blast amputation model (93). In this model extremity amputation was produced through detonation of an explosive while

protecting the animal proximal to the specified amputation level. This model was able to produce HO with a good reliability (4/4 hind limb, 1/5 fore limb) especially in hind limb amputations.

Based on the presented reports, we conclude that most of the models described here do not seem to be suffciently reliable to be routinely used. It remains unclear whether the formation of bone as described in these studies can be defined as HO. The recent model of Tannous *et al* might be of relevance to study the formation of HO after blast amputation in war setting, as HO in the residual limbs of combat-related amputees has been reported in up to 63% of patients (71,93).

Irritant injection model

Heinen et al reported the induction of HO in rabbits by injection of 40 % ethanol (35). As the injection of various irritant substances into muscles was reported to lead to the formation of HO, many trials to identify further substances inducing HO were undertaken. Selle and Urist for example reported that acid-alcohol could induce HO in a small percent of animals, while injections of calcium chloride led to calcification in soft tissue only (85). Others used a mixture of phosphatase glycerophosphate and alginate gel (7). In contrast, in their search to find a specific osteogenetic substance, alcoholic subtracts of bone were used. In most of these publications, pure alcohol is used as a control, but no HO formation was found (3,49,50,73). The insufficient repeatability and questionable clinical relevance of these models grossly limits their potential use.

CONCLUSION

Currently 6 animal models are known to mimic HO. Most enable to reflect some forms, particular aspects, or only distinct varieties of the human condition. The questionable reliability and ambiguous clinical relevance of most of the models complicate their use in the search for prophylactic and therapeutic treatments for HO. Results of studies with models with unknown relevance to the condition in human should therefore be carefully examined, as conclusions and interpolation of the respective results may not always be comparable to the human condition. It is therefore the authors opinion that, in order to get a full understanding of the pathogenesis of HO, more mechanism-based models as the hip-surgery model and the extremity blast amputation model are needed. In the mean time, it remains important to choose the right model that fits the question asked in the study of HO and be aware of its limitations when drawing conclusions towards the human condition.

REFERENCES

- Ahn J, Serrano de la Pena L, Shore EM, Kaplan FS. Paresis of a bone morphogenetic protein-antagonist response in a genetic disorder of heterotopic skeletogenesis. J. Bone Joint Surg Am 2003; 85-A: 667-674.
- Ahrengart L. Periarticular heterotopic ossification after total hip arthroplasty. Risk factors and consequences. *Clin Orthop* 1991; 49-58.
- **3.** Annersten S. Experimentelle untersuchungen über die Osteogenese und die Biochemie des Fracturcallus. *Acta Chir Scand* 1940 ; Supp 60.
- 4. Baird EO, Kang QK. Prophylaxis of heterotopic ossification - an updated review. J Orthop Surg 2009 ; 4 : 12.
- 5. Bartlett CS, Rapuano BE, Lorich DG, Wu T, Anderson RC, Tomin E *et al.* Early changes in prostaglandins precede bone formation in a rabbit model of heterotopic ossification, *Bone* 2006; 38 : 322-332.
- Bertelsen A. Experimental Investigations into Post-Foetal Osteogenesis. Acta Chir. Scand 1944; 139-181.
- 7. Blum G. Phosphatase and the repair of fractures. *The Lancet* 1994; 75-78.
- **8.** Brooker AF, Bowerman JW, Robinson RA, Riley Jr LH. Ectopic ossification following total hip replacement. Incidence and a method of classification. *J. Bone Joint Surg Am* 1973 ; 55 ; 1629-1632.
- **9.** Buck RC. Regeneration of tendon, *J. Pathol. Bacteriol.* 1953 ; 66 : 1-18.
- 10. Burwell RG. Studies in the transplantation od Bone. The Fresh Composite Homograft-Autograft of Cancellous Bone; an Analysis of Factors Leading to Osteogenesis in Marrow Transplants and in Marrow-Containing Bone Grafts. J. Bone Joint Surg Br 1964; 46 : 110-140.
- Chalmers J, Gray DH, Rush J. Observations on the induction of bone in soft tissues. J. Bone Joint Surg Br: 1975; 57: 36-45.
- 12. Circi E, Akpinar S, Balcik C, Bacanli D, Guven G, Akgun RG et al. Biomechanical and histological comparison of the influence of oestrogen deficient state on tendon healing potential in rats. Int. Orthop. 2009; 33: 1461-1466.

- **13. Collins M.** Experimental myositis ossificans in dogs. *J Bone Jt Surg* 1965 ; 47 : 1277.
- 14. Craven PL, Urist MR. Osteogenesis by radioisotope labelled cell populations in implants of bone matrix under the influence of ionizing radiation. *Clin Orthop* 1971; 76: 231-243.
- **15.** Danis A. [Study of ossification in bone marrow grafts], *Acta Chir Belg* 1957 ; 56 : 1-120.
- Dejerine A, Ceillier A. Para-osteo-arthropathies des paraplegigues par lesion medullarie; etude clinique et radiographique. *Ann Med* 1918; 5: 497.
- DiCesare PE, Nimni ME, Peng L, Yazdi M, Cheung DT. Effects of indomethacin on demineralized boneinduced heterotopic ossification in the rat. J Orthop Res Off Publ Orthop Res Soc 1991; 9: 855-861.
- Ekelund A, Brosjö O, Nilsson OS. Experimental induction of heterotopic bone, *Clin Orthop* 1991; 102-112.
- **19.** Friedenstein AJ, Piatetzky-Shapiro II, Petrakova KV. Osteogenesis in transplants of bone marrow cells. *J Embryol Exp Morphol* 1966; 16: 381-390.
- Garland DE. A clinical perspective on common forms of acquired heterotopic ossification. *Clin Orthop* 1991; 13-29.
- Garland DE, Blum CE, Waters RL. Periarticular heterotopic ossification in head-injured adults. Incidence and location. J Bone Joint Surg Am 1980; 62: 1143-1146.
- 22. Garland DE, O'Hollaren RM. Fractures and dislocations about the elbow in the head-injured adult, *Clin Orthop* 1982; 38-41.
- 23. Gazzerro E, Gangji V, Canalis E. Bone morphogenetic proteins induce the expression of noggin, which limits their activity in cultured rat osteoblasts. *J Clin Invest* 1998; 102: 2106-2114.
- 24. Geschikter CF, Maseritz IH. Myositis Ossificans. *J Bone Jt Surg* 1938 ; 20 : 66-674.
- 25. Glaser DL, Economides DN, Wang L, Liu X, Kimble RD, Fandl JP *et al.* In vivo somatic cell gene transfer of an engineered Noggin mutein prevents BMP4-induced heterotopic ossification. *J Bone Joint Surg Am* 2003; 85-A : 2332-2342.
- 26. Gonda K, Nakaoka T, Yoshimura K, Otawara-Hamamoto Y, Harrii K. Heterotopic ossification of degenerating rat skeletal muscle induced by adenovirusmediated transfer of bone morphogenetic protein-2 gene. *J Bone Miner Res Off J Am Soc Bone Miner Res* 2000; 15: 1056-1065.
- 27. Griffin SM, Sims SH, Karunakar MA, Seymour R, Haines N. Heterotopic Ossification Rates After Acetabular Fracture Surgery Are Unchanged Without Indomethacin Prophylaxis. *Clin Orthop* 2013; Feb 26. [Epub ahead of print]
- Gruber GB. Über Histologie und Pathogenese der Circumskripter Muskelverknocherung. Fischer, Jena, 1913.
- 29. Haga F, Fujimura A. Über Myositis ossificans traumatica (reit- u. Exerzierknochen). *Arch Klin Chir* 1904 ; 64.

- **30. Hait G, Boswick Jr JA, Stone NH.** Heterotopic bone formation secondary to trauma (myositis ossificans traumatica). *J Trauma* 1970; 10: 405-411.
- 31. Hannallah D, Peng H, Young B, Usas A, Gearhart B, Huard J. Retroviral delivery of Noggin inhibits the formation of heterotopic ossification induced by BMP-4, demineralized bone matrix, and trauma in an animal model. J Bone Joint Surg Am 2004; 86-A : 80-91.
- 32. Haque S, Eisen RN, West AB. Heterotopic bone formation in the gastrointestinal tract. *Arch Pathol Lab Med* 1996; 120: 666-670.
- **33. Hardy JR, Rooney P.** Use of the myositis ossificans model of Michelsson. *Clin Orthop* 1997 : 340-342.
- 34. Hayashi C, Hasegawa U, Saita Y, Hemmi H, Hayata T, Nakashima K et al. Osteoblastic bone formation is induced by using nanogel-crosslinking hydrogel as novel scaffold for bone growth factor. J Cell Physiol 2009; 220; 1-7.
- **35.** Heinen JH Jr, Dabbs GH, Mason HA. The experimental production of ectopic cartilage and bone in the muscles of rabbits. *J Bone Joint Surg Am* 1949; 31A : 765-775.
- **36.** Horne LT, Blue BA. Intraarticular heterotopic ossification in the knee following intramedullary nailing of the fractured femur using a retrograde method, *J Orthop Trauma* 1999; 13: 385-388.
- 37. Jacobs JE, Birnbaum BA, Siegelman ES. Heterotopic ossification of midline abdominal incisions : CT and MR imaging findings. *Ajr Am J Roentgenol* 1996; 166: 579-584.
- **38. Jensen LL, Halar E, Little JW, Brooke MM.** Neurogenic heterotopic ossification. *Am J Phys Med* 1987; 66: 351-363.
- **39.** Jones RW. Ossification of the Achilles Tendon. BMJ 1932; 2: 943-943.
- Kan L, Kessler JA. Animal models of typical heterotopic ossification. J Biomed Biotechnol 2011; 2011: 309287.
- 41. Kan L, Liu Y, McGuire TL, Berger DMP, Awatramani RB, Dymecki SM et al. Dysregulation of local stem/progenitor cells as a common cellular mechanism for heterotopic ossification. Stem Cells Dayt Ohio 2009; 27: 150-156.
- 42. Kan L, Lounev VY, Pignolo RJ, Duan L, Liu Y, Stock RS et al. Substance P signaling mediates BMPdependent heterotopic ossification, J Cell Biochem 2011; 112 : 2759-2772.
- 43. Kaplan FS, Smith RM. Fibrodysplasia ossificans progressiva (FOP). J Bone Miner Res Off J Am Soc Bone Miner Res 1997; 12: 855.
- **44. van Kuijk AA, Geurts ACH, van Kuppevelt HJM.** Neurogenic heterotopic ossification in spinal cord injury. *Spinal Cord* 2002 ; 40 : 313-326.
- **45.** Lacroix P. Recent Investigations on the growth of bone. *Nature* 194 ; 156.
- 46. Lagos MF, Zarapico RM. Obtension experimental de hueso metaplasico. Inst Nac Cien Med 1946; 173-215.

- 47. Leblanc E, Trensz F, Haroun S, Drouin G, Bergeron E, Penton CM et al. BMP-9-induced muscle heterotopic ossification requires changes to the skeletal muscle microenvironment. J Bone Miner Res Off J Am Soc Bone Miner Res 2011; 26: 1166-1177.
- **48.** Levander G. A Study of bone regeneration. *Surg Gynecol Obst* 1938.
- 49. Levander G. Tissue Induction. Nature 1945 ; 148-149.
- **50.** Levander G, Willstaedt H. Alcohol-soluble osteogenetic substance from bone marrow. *Nature* 1946 ; 157 : 587.
- 51. Lin L, Shen Q, Leng H, Duan X, Fu X, Yu C. Synergistic inhibition of endochondral bone formation by silencing Hifl α and Runx2 in trauma-induced heterotopic ossification. *Mol Ther J Am Soc Gene Ther* 2011; 19: 1426-1432.
- 52. Lin L, Shen Q, Xue T, Yu C. Heterotopic ossification induced by Achilles tenotomy via endochondral bone formation : expression of bone and cartilage related genes. *Bone* 2010 ; 46 : 425-431.
- **53.** Linan E, O'Dell MW, Pierce JM. Continuous passive motion in the management of heterotopic ossification in a brain injured patient. *Am J Phys Med Rehabil Assoc Acad Physiatr* 2001; 80 : 614-617.
- 54. Lindholm TC, Lindholm TS, Nilsson OS, Sjökvist G. Influence of 1 alpha-hydroxyvitamin D3 and 24,25dihydroxyvitamin D3 on experimentally induced heterotopic ossification in rats, Scand. *J Rheumatol* 1986; 15: 68-74.
- 55. Lounev VY, Ramachandran R, Wosczyna MN, Yamamoto M, Maidment ADA, Shore EM et al. Identification of progenitor cells that contribute to heterotopic skeletogenesis. J Bone Joint Surg Am 2009; 91: 652-663.
- **56.** McCarthy EF, Sundaram M. Heterotopic ossification : a review. *Skeletal Radiol* 2005; 34 : 609-619.
- McClure J. The effect of diphosphonates on heterotopic ossification in regenerating Achilles tendon of the mouse. *J Pathol* 1983 ; 139 : 419-430.
- 58. Michelsson JE, Granroth G, Andersson LC. Myositis ossificans following forcible manipulation of the leg. A rabbit model for the study of heterotopic bone formation. *J Bone Joint Surg Am* 1980; 62: 811-815.
- **59.** Michelsson JE, Pettilä M, Valtakari T, Leivo I, Aho HJ. Isolation of bone from muscles prevents the development of experimental callus-like heterotopic bone. A study of the interaction of bone and muscle in new bone formation. *Clin Orthop* 1994; 266-272.
- Moed BR, Resnick RB, Fakhouri AJ, Nallamothu B, Wagner RA. Effect of two nonsteroidal antiinflammatory drugs on heterotopic bone formation in a rabbit model. J Arthroplasty 1994; 9: 81-87.
- 61. Murat N, Hocaoglu N, Karatosun V, Yorukoglu K, Gidener S, Gunal I. The effects of non-selective and cyclooxygenase-2-selective non-steroidal anti-inflammatory drugs on heterotopic ossification in rats. *Med Sci Monit Int Med J Exp Clin Res* 2005; 11: BR449-451.

8

- Namazi H, Mozaffarian K. Levothyroxin inhibits heterotopic ossification: an experimental study in rabbits *J Trauma* 2008; 65: 849-851.
- 63. Le Nihouannen D, Daculsi G, Saffarzadeh A, Gauthier O, Delplace S, Pilet P et al. Ectopic bone formation by microporous calcium phosphate ceramic particles in sheep muscles. *Bone* 2005; 36: 1086-1093.
- O'Connor JP. Animal models of heterotopic ossification. Clin. Orthop. 1998; 71-80.
- **65.** Oh MM, Kim JJ, Kang SH, Park HS, Moon DG, Bae JH. Heterotrophic bone formation with bone marrow in the kidney parenchyme. *Urol Res* 2010; 38: 409-410.
- 66. Osawa K, Okubo Y, Nakao K, Koyama N, Bessho K. Osteoinduction by microbubble-enhanced transcutaneous sonoporation of human bone morphogenetic protein-2. *J Gene Med* 2009; 11: 633-641.
- 67. Pape HC, Marsh S, Morley JR, Krettek C, Giannoudis PV. Current concepts in the development of heterotopic ossification. J Bone Joint Surg Br 2004; 86 : 783-787.
- 68. Patin G, Tome I, Du Laurens P. Lettres choisies de feu M. Guy Patin dans lesquelles sont contenués plusieurs particularités historiques sur la vie et la mort des savans de ce siècle, sur leurs écrits, et sur plusieurs autres choses curieuses depuis l'an 1645 jusqu'en 1672. Cologne. 1692, 28.
- 69. Peng H, Chen ST, Wergedal JE, Polo JM, Yee JK, Lau KH et al. Development of an MFG-based retroviral vector system for secretion of high levels of functionally active human BMP4. *Mol Ther J Am Soc Gene Ther* 2001; 4: 95-104.
- 70. Peng H, Wright V, Usas A, Gearhart B, Shen HC, Cummins J et al. Synergistic enhancement of bone formation and healing by stem cell-expressed VEGF and bone morphogenetic protein-4. J Clin Invest 2002; 110: 751-759.
- Potter BK, Burns TC, Lacap AP, Granville RR, Gajewski DA. Heterotopic ossification following traumatic and combat-related amputations. Prevalence, risk factors, and preliminary results of excision. *J Bone Joint* Surg Am 2007; 89: 476-486.
- 72. Reardon MJ, Tillou A, Mody DR, Reardon PR. Heterotopic calcification in abdominal wounds. Am J Surg 1997; 173: 145-147.
- Redano C. Su di un particolare principio osteogenetico contenuto nelle ossa normali. *Ann Ital Chir* 1942; 249-271.
- Reddi AH, Huggins C. Biochemical sequences in the transformation of normal fibroblasts in adolescent rats. *Proc. Natl. Acad. Sci. U. S. A.* 1972; 69: 1601-1605.
- **75. Riedel B.** Demonstration line durch ach Hagiges Umhergehen total destruirten kniegelenkes von einem patienten mit stichverletzing des ruckans. *Verh Dtsch Ges Chir* 1883; 12:93.

- **76. Ritter MA, Sieber JM.** Prophylactic indomethacin for the prevention of heterotopic bone formation following total hip arthroplasty. *Clin. Orthop.* 1985 : 217-225.
- Ritter MA, Vaughan RB. Ectopic ossification after total hip arthroplasty. Predisposing factors, frequency, and effect on results. J. Bone Joint Surg. Am. 1977; 59: 345-351.
- 78. Rosendahl S, Christoffersen JK, Norgaard M. Paraarticular ossification following hip replacement. 70 arthroplasties ad modum Moore using McFarland's approach. *Acta Orthop Scand* 1977; 48: 400-404.
- 79. Rumi MN, Deol GS, Bergandi JA, Singapuri KP, Pellegrini VD Jr. Optimal timing of preoperative radiation for prophylaxis against heterotopic ossification. A rabbit hip model. J Bone Joint Surg Am 2005; 87: 366-373.
- 80. Rumi MN, Deol GS, Singapuri KP, Pellegrini VD Jr. The origin of osteoprogenitor cells responsible for heterotopic ossification following hip surgery: an animal model in the rabbit. J Orthop Res Off Publ Orthop Res Soc 2005; 23: 34-40.
- **81. Salah ED.** Heterotopic ossification of the tendo achillis. *J Anat* 1967 ; 101 : 611.
- Salah ED, Pritchard JJ. Heterotopic ossification in the tendo achillis of the rat following crushing and ligation. *J Anat* 1969; 104: 181.
- 83. Sandberg O, Eliasson P, Andersson T, Agholme F, Aspenberg P. Etanercept does not impair healing in rat models of tendon or metaphyseal bone injury. *Acta Orthop* 2012; 83: 305-310.
- 84. Schneider DJ, Moulton MJ, Singapuri K, Chinchilli V, Deol GS, Krenitsky G et al. The Frank Stinchfield Award. Inhibition of heterotopic ossification with radiation therapy in an animal model. Clin Orthop 1998; 35-46.
- **85. Selle RW, Urist MR.** Calcium deposits and new bone formation in muscle in rabbits. *J Surg Res* 1961; 1:132-141.
- 86. Shafritz AB, Shore EM, Gannon FH, Zasloff MA, Taub R, Muenke M et al. Overexpression of an osteogenic morphogen in fibrodysplasia ossificans progressiva. N Engl J Med 1996; 335: 555-561.
- Shehab D, Elgazzar AH, Collier BD. Heterotopic ossification. J. Nucl. Med. Off. Publ. Soc. Nucl. Med. 2002; 43: 346-353.
- 88. Shimono K, Morrison TN, Tung W, Chandraratna RA, Williams JA, Iwamoto M et al. Inhibition of ectopic bone formation by a selective retinoic acid receptor alpha-agonist : a new therapy for heterotopic ossification? J Orthop Res Off Publ Orthop Res Soc 2010; 28 : 271-277.
- 89. Shimono K, Tung WE, Macolino C, Chi AHT, Didizian JH, Mundy C *et al.* Potent inhibition of heterotopic ossification by nuclear retinoic acid receptor-γ agonists. *Nat Med* 2011; 17: 454-460.
- 90. Shore EM, Gannon FH, Kaplan FS. Fibrodysplasia ossificans progressiva why do some people have two skeletons? J Clin Rheumatol Pr Reports Rheum Musculoskelet Dis 1997; 3: 84-89.

- 91. Stone CA Ossifying Hematoma. JAMA 1926; 1885.
- **92.** Stover SL, Niemann KM, Tulloss JR. Experience with surgical resection of heterotopic bone in spinal cord injury patients. *Clin Orthop* 1991 : 71-77.
- **93. Tannous O, Griffith C, O'Toole RV, Pellegrini VD Jr.** Heterotopic ossification after extremity blast amputation in a Sprague-Dawley rat animal model. *J Orthop Trauma* 2011; 25 : 506-510.
- 94. Tannous O, Stall AC, Griffith C, Donaldson CT, Castellani RJ Jr, Pellegrini VD Jr. Heterotopic bone formation about the hip undergoes endochondral ossification: a rabbit model. *Clin Orthop* 2013; 471: 1584-1592.
- **95. Thomas BJ.** Heterotopic bone formation after total hip arthroplasty. *Orthop Clin North Am* 1992; 23: 347-358.
- **96.** Toom A, Suutre S, Märtson A, Haviko T, Selstam G, Arend A. Lack of a central role for osteoprogenitor cells from the femoral canal in heterotopic ossification of the hip : an experimental study in a rat model. *J Bone Joint Surg Br* 2010 ; 92 : 298-303.
- **97. Törnkvist H, Nilsson OS, Bauer FC, Lindholm TS.** Experimentally induced heterotopic ossification in rats influenced by anti-inflammatory drugs. *Scand J Rheumatol* 1983; 12: 177-180.
- 8. Tsailas PG, Babis GC, Nikolopoulos K, Soucacos PN, Korres DS. The effectiveness of two COX-2 inhibitors in the prophylaxis against heterotopic new bone formation : an experimental study in rabbits. J. Surg. Res. 2009; 151; 108-114.
- 99. Tsumaki N, Nakase T, Miyaji T, Kakiuchi M, Kimura T, Ochi T et al. Bone morphogenetic protein signals are required for cartilage formation and differently regulate joint development during skeletogenesis, J Bone Miner Res Off J Am Soc Bone Miner Res 2002; 17: 898-906.
- **100.** Urist MR. Bone: formation by autoinduction. *Science* 1965; 150: 893-899.
- 101. Urist MR, Mikulski A, Lietze A. Solubilized and insolubilized bone morphogenetic protein. *Proc Natl Acad Sci USA* 1979; 76: 1828-1832.
- **102.** Urist MR, Mikulski AJ. A soluble bone morphogenetic protein extracted from bone matrix with a mixed aqueous and nonaqueous solvent. *Proc Soc Exp Biol Med Soc Exp Biol Med New York N* 1979 ; 162 : 48-53.
- 103. Vanden Bossche L, Vanderstraeten G. Heterotopic ossification : a review. J Rehabil Med Off J Uems Eur Board Phys Rehabil Med 2005 ; 37 : 129-136.
- 104. Vanden Bossche LC, Van Maele G, Wojtowicz I, Bru I, Decorte T, De Muynck M et al. Free radical scavengers versus methylprednisolone in the prevention of experimentally induced heterotopic ossification. J Orthop Res Off Publ Orthop Res Soc 2009; 27: 748-751.
- 105. Vanden Bossche LC, Van Maele G, Wojtowicz I, De Cock K, Vertriest S, De Muynck M *et al.* Free radical

scavengers are more effective than indomethacin in the prevention of experimentally induced heterotopic ossification. *J Orthop Res Off Publ Orthop Res Soc* 2007; 25: 267-272.

- 106. Volek-Smith H, Urist MR. Recombinant human bone morphogenetic protein (rhBMP) induced heterotopic bone development in vivo and in vitro. Proc Soc Exp Biol Med Soc Exp Biol Med New York N 1996; 211: 265-272.
- 107. Walton M, Rothwell AG. Reactions of thigh tissues of sheep to blunt trauma. *Clin Orthop* 1983; 273-281.
- 108. Wozney JM, Rosen V, Celeste AJ, Mitsock LM, Whitters MJ, Kriz RW, et al. Novel regulators of bone formation : molecular clones and activities. *Science* 1988 ; 242 : 1528-1534.
- 109. Wright V, Peng H, Usas A, Young B, Gearhart B, Cummins J, et al. BMP4-expressing muscle-derived stem cells differentiate into osteogenic lineage and improve bone healing in immunocompetent mice. Mol Ther J Am Soc Gene Ther 2002; 6: 169-178.
- 110. Yamamoto K, Hojo H, Koshima I, Chung U, Ohba S. Famotidine suppresses osteogenic differentiation of tendon cells in vitro and pathological calcification of tendon in vivo. J Orthop Res Off Publ Orthop Res Soc 2012; 30: 1958-1962.
- 111. Yu PB, Deng DY, Lai CS, Hong CC, Cuny GD, Bouxsein ML *et al.* BMP type I receptor inhibition reduces heterotopic [corrected] ossification. *Nat Med* 2008; 14: 1363-1369.
- 112. Yuan H, De Bruijn JD, Li Y, Feng J, Yang Z, De Groot K, *et al.* Bone formation induced by calcium phosphate ceramics in soft tissue of dogs : a comparative study between porous alpha-TCP and beta-TCP. *J Mater Sci Mater Med* 2001 ; 12 : 7-13.
- **113. Zaccalini PS, Urist MR.** Traumatic Periosteal Proliferations in Rabbits. The Enigma of Experimental Myositis Ossificans Traumatica. *J Trauma* 1964; 4: 344-357.
- 114. Zhang K, Wang L, Zhang S, Yu B, Liu F, Cui Z et al. Celecoxib inhibits the heterotopic ossification in the rat model with Achilles tenotomy. Eur J Orthop Surg Traumatol Orthopédie Traumatol 2013; 23: 145-148.
- **115. Zimmerman LB, De Jesús-Escobar JM, Harland RM** The Spemann organizer signal noggin binds and inactivates bone morphogenetic protein 4. *Cell* 1996 ; 86 : 599-606.
- 116. Zimmermann SM, Würgler-Hauri CC, Wanner GA, Simmen HP, Werner CML Echinomycin in the prevention of heterotopic ossification – an experimental antibiotic agent shows promising results in a murine model. *Injury* 2013 ; 44 : 570-575.
- 117. Zotz TGG, de Paula JB, Moser ADL Experimental model of heterotopic ossification in Wistar rats. Braz J Med Biol Res Rev Bras Pesqui Médicas E Biológicas Soc Bras Biofísica Al 2012; 45: 497-501.